

The Proton Pump Inhibitor Omeprazole Does Not Promote *Clostridioides difficile* Colonization in a Murine Model

Sarah Tomkovich,^a Nicholas A. Lesniak,^a Yuan Li,^a Lucas Bishop,^a Madison J. Fitzgerald,^a  Patrick D. Schloss^a

^aDepartment of Microbiology and Immunology, University of Michigan, Ann Arbor, Michigan, USA

ABSTRACT Proton pump inhibitor (PPI) use has been associated with microbiota alterations and susceptibility to *Clostridioides difficile* infections (CDIs) in humans. We assessed how PPI treatment alters the fecal microbiota and whether treatment promotes CDIs in a mouse model. Mice receiving a PPI treatment were gavaged with 40 mg of omeprazole per kg of body weight during a 7-day pretreatment phase, the day of *C. difficile* challenge, and the following 9 days. We found that mice treated with omeprazole were not colonized by *C. difficile*. When omeprazole treatment was combined with a single clindamycin treatment, one cage of mice remained resistant to *C. difficile* colonization, while the other cage was colonized. Treating mice with only clindamycin followed by challenge resulted in *C. difficile* colonization. 16S rRNA gene sequencing analysis revealed that omeprazole had minimal impact on the structure of the murine microbiota throughout the 16 days of omeprazole exposure. These results suggest that omeprazole treatment alone is not sufficient to disrupt microbiota resistance to *C. difficile* infection in mice that are normally resistant in the absence of antibiotic treatment.

IMPORTANCE Antibiotics are the primary risk factor for *Clostridioides difficile* infections (CDIs), but other factors may also increase a person's risk. In epidemiological studies, proton pump inhibitor (PPI) use has been associated with CDI incidence and recurrence. PPIs have also been associated with alterations in the human intestinal microbiota in observational and interventional studies. We evaluated the effects of the PPI omeprazole on the structure of the murine intestinal microbiota and its ability to disrupt colonization resistance to *C. difficile*. We found omeprazole treatment had minimal impact on the murine fecal microbiota and did not promote *C. difficile* colonization. Further studies are needed to determine whether other factors contribute to the association between PPIs and CDIs seen in humans or whether aspects of murine physiology may limit its utility to test these types of hypotheses.

KEYWORDS 16S rRNA gene, *Clostridioides difficile*, *Clostridium difficile*, colonization resistance, infection, microbial ecology, microbiome, mouse model, pathogenesis


Antibiotics have a large impact on the intestinal microbiome and are a primary risk factor for developing *Clostridioides difficile* infections (CDIs) (1). It is less clear whether other human medications that impact the microbiota also influence *C. difficile* colonization resistance. The results of multiple epidemiological studies have suggested an association between proton pump inhibitor (PPI) use and incidence or recurrence of CDIs (2–5). There have also been a number of large cohort studies and interventional clinical trials that demonstrated that specific alterations in the intestinal microbiome were associated with PPI use (4, 6). PPI-associated microbiota changes have been attributed to the ability of PPIs to increase stomach acid pH which may promote the survival of oral and pathogenic bacteria (4, 6). In human fecal samples, PPI use results in increases in *Enterococcaceae*, *Lactobacillaceae*, *Micrococcaceae*, *Staphylococcaceae*,

Citation Tomkovich S, Lesniak NA, Li Y, Bishop L, Fitzgerald MJ, Schloss PD. 2019. The proton pump inhibitor omeprazole does not promote *Clostridioides difficile* colonization in a murine model. mSphere 4:e00693-19. <https://doi.org/10.1128/mSphere.00693-19>.

Editor Patricia A. Bradford, Antimicrobial Development Specialists, LLC

Copyright © 2019 Tomkovich et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Patrick D. Schloss, pschloss@umich.edu.

 PPI (omeprazole) treatment alone is not sufficient to disrupt colonization resistance to *C. difficile* infection in mice that are normally resistant in the absence of antibiotic treatment. @PatSchloss

Received 17 September 2019

Accepted 6 November 2019

Published 20 November 2019

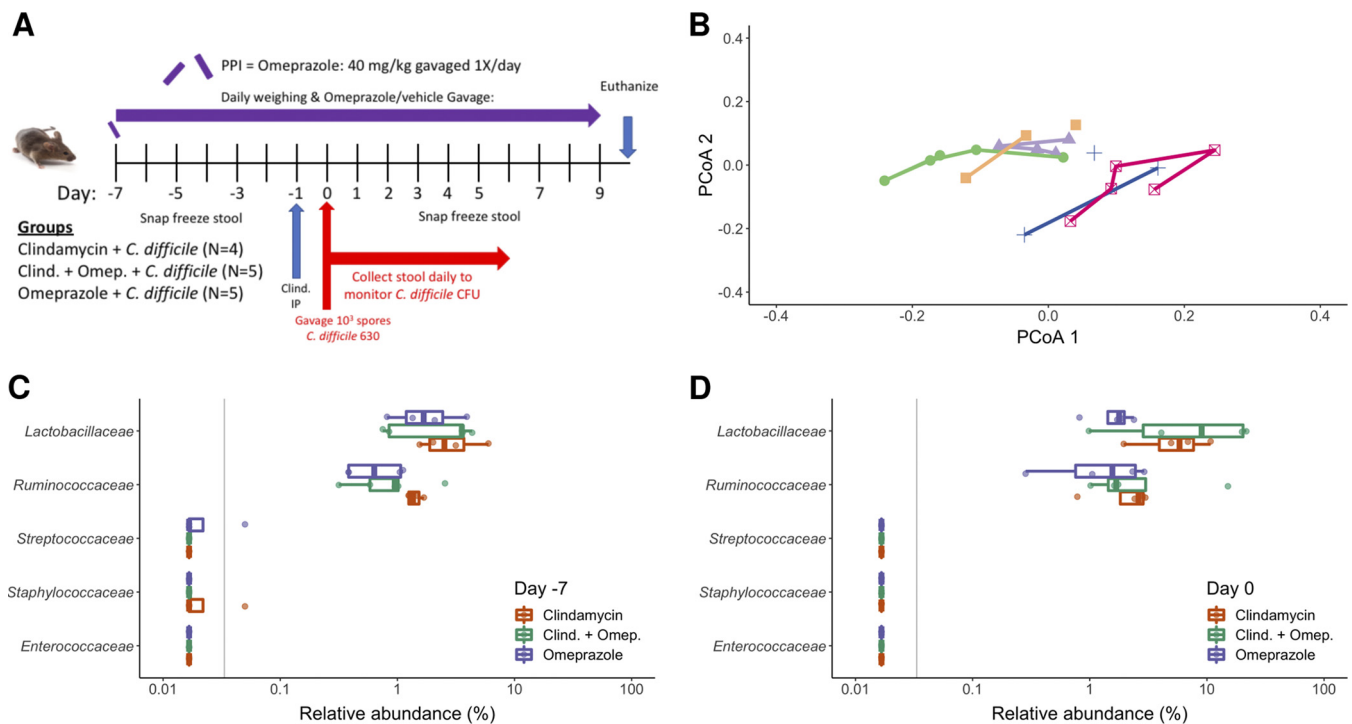


FIG 1 Omeprazole treatment had minimal impact on the murine fecal microbiota. (A) Mouse experiment timeline and logistics. The PPI omeprazole was administered throughout the duration of the experiment. Clindamycin was administered intraperitoneally (IP) 1 day before *C. difficile* challenge on day 0. Stool samples for 16S rRNA sequencing analysis were collected on the days that are labeled (days -7, -5, -3, -1, 0, 1, 2, 3, 4, 5, 7, and 9). *C. difficile* CFU in the stool was quantified daily through 6 days postinfection by anaerobic culture. (B) Principal coordinate analysis (PCoA) of Bray-Curtis distances from stool samples of mice in the omeprazole treatment group during the initial 7 days of the experiment. Each color represents stool samples from the same mouse, and lines connect sequentially collected samples. (C and D) Relative abundances of families previously associated with PPI use in humans at the start of the experiment (C) and after 7 days of omeprazole treatment (D). Each circle represents the value for an individual mouse. There were no significant differences across treatment groups for any of the identified families in the sequence data at day -7 (all P values > 0.448) and day 0 (all P values > 0.137), analyzed by Kruskal-Wallis test with a Benjamini-Hochberg correction for multiple comparisons. In panels C and D, the gray vertical line indicates the limit of detection.

and *Streptococcaceae* and decreases in *Ruminococcaceae* (6–9). Several of these taxa have also been associated with *C. difficile* colonization in humans (10).

Unfortunately, the studies suggesting a link between PPIs and *C. difficile* were retrospective and did not evaluate changes in the microbiome (2, 3, 5). Thus, it is unclear whether the gastrointestinal microbiome changes associated with PPI use explain the association between PPIs and CDIs. Additionally, epidemiological studies have a limited capacity to address potential confounders and comorbidities in patients who were on PPIs and developed CDIs or recurrent CDIs (2, 5). Here, we evaluated the impact of daily PPI treatment with omeprazole on the murine microbiome and susceptibility to *C. difficile* colonization in relation to clindamycin, an antibiotic that perturbs the microbiome enough to allow *C. difficile* to colonize but is mild enough that *C. difficile* is cleared within 10 days (11).

Murine fecal microbiomes were minimally affected by omeprazole treatment.

To test whether omeprazole treatment alters the microbiome and promotes susceptibility to CDIs, we gavaged mice with 40 mg of omeprazole per kg of body weight for 7 days before *C. difficile* challenge (Fig. 1A). A principal coordinate analysis (PCoA) of the Bray-Curtis distances over the initial 7 days of treatment revealed that the bacterial communities of omeprazole-treated mice remained relatively unchanged (Fig. 1B). We observed no significant changes in the relative abundance of those taxa previously shown to respond to PPI treatment throughout the course of the 16-day experiment (Fig. 1C and D; see also Fig. S1 in the supplemental material). We also observed no significant changes in relative abundances at the family and genus level over the course of the experiment for the omeprazole-treated mice (all corrected P values > 0.36). These results demonstrated that the omeprazole treatment alone had minimal impact on the murine fecal bacterial community after 7 days of pretreatment.

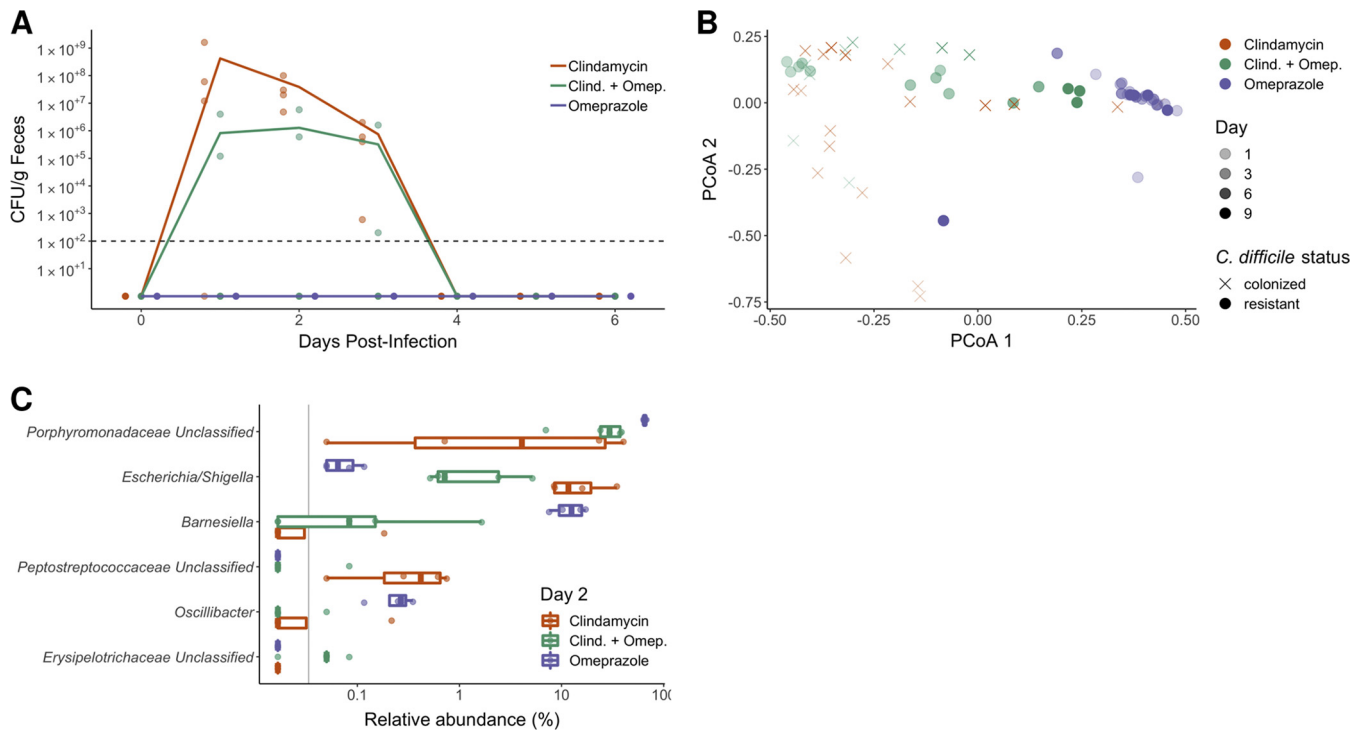


FIG 2 Omeprazole treatment alone does not promote CDIs in mice. (A) *C. difficile* CFU/gram of stool measured each day after *C. difficile* challenge for mice treated with clindamycin, clindamycin plus omeprazole, and omeprazole. The lines represent the mean CFU/gram for each treatment group, while points represent CFU/gram for individual mice within each group. The black dashed line indicates the limit of detection. (B) PCoA of Bray-Curtis distances from stool samples collected after antibiotic treatment (last 9 days of the experiment). The hue of the symbol indicates the treatment day. Symbols represent the *C. difficile* colonization status of the mice measured 2 days postinfection. Circles represent resistant mice (*C. difficile* was undetectable in stool samples), while × symbols represent mice that were colonized with *C. difficile*, although all mice cleared *C. difficile* within 5 days of infection. Omeprazole-treated fecal samples primarily cluster together throughout the experiment. (C) Genera that vary the most across treatment groups for stool samples collected from mice 2 days postinfection. Data were analyzed by Kruskal-Wallis test, and no *P* values were significant after Benjamini-Hochberg correction for multiple comparisons (all *P* values > 0.092). The gray vertical line indicates the limit of detection.

Omeprazole treatment did not promote susceptibility to *C. difficile* infection in mice. Next, we examined whether omeprazole treatment altered susceptibility to *C. difficile* infection in mice. After omeprazole or clindamycin treatment, mice were challenged with 10^3 *C. difficile* strain 630 spores. Although *C. difficile* colonized the clindamycin-treated mice, it did not colonize the omeprazole-treated mice (Fig. 2A). Interestingly, only one cage of mice that received both omeprazole and clindamycin were colonized, while the other cage of mice were resistant (Fig. 2A). The greatest shifts in bacterial communities occurred in the clindamycin-treated mice (Fig. 2B and Fig. S2). Regardless of whether the mice were colonized by *C. difficile*, all of the mice had cleared *C. difficile* within 5 days (Fig. 2A), suggesting that omeprazole did not affect the rate of clearance. Our results suggest that omeprazole treatment had no effect on bacterial community resistance to *C. difficile* colonization in mice. Instead, most of the differences between the three treatment groups appeared to be driven by clindamycin administration (Fig. 2C and Fig. S2). These findings demonstrated that high-dose omeprazole treatment did not promote susceptibility to *C. difficile* colonization.

Conclusions. The PPI omeprazole did not meaningfully impact the structure of the gut microbiota and did not promote *C. difficile* infection in mice. Our findings that omeprazole treatment had minimal impact on the fecal microbiome were comparable to another PPI mouse study that indicated the PPI lansoprazole had more of an effect on the small intestinal microbiota compared to the fecal microbiota (12). The same group demonstrated lansoprazole treatment increased the stomach pH in mice (12), which may improve survival of bacteria passing through the stomach. We did not find significant changes in the relative abundances of the taxa observed to be significantly impacted by PPI use in human studies. However, three of the human-associated taxa

were absent or at low abundance in our mice. Interestingly, other groups examining fecal microbiota communities before and after PPI administration to healthy cats and infants with gastroesophageal reflux disease found that PPIs have minimal effects on fecal bacterial community structures, although there were a few significant changes in specific genera (13, 14). One limitation of our study is that there were only four or five mice per group, which may have limited our ability to identify PPI-induced changes in specific bacterial genera. Although our fecal microbiota findings are comparable to what has been shown in another mouse study (12), whether PPI-induced changes in specific bacterial abundances observed in humans play a role in CDIs remains to be determined.

Although several *C. difficile* mouse model studies have shown that PPIs have an effect on CDIs with or without additional antibiotic treatment (15–17), there were insufficient controls to attribute the effect solely to PPI treatment. One group administered 0.5 mg/kg of the PPI lansoprazole daily for 2 weeks to mice and then challenged with *C. difficile* demonstrated that PPI treatment alone resulted in detectable *C. difficile* in stools 1 week after challenge; however, there was detectable *C. difficile* in mice not treated with antibiotics (15, 16). The other mouse study demonstrated that antibiotic- and esomeprazole-treated mice developed more severe CDIs than antibiotic-treated mice, but the researchers did not have a group treated with just esomeprazole for comparison (17). We tested the same high 40-mg/kg PPI dose and expanded pretreatment to 7 days before challenge to test the impact of omeprazole treatment alone on our CDI mouse model. Additionally, we have previously demonstrated that mice from our breeding colony are resistant to *C. difficile* 630 colonization without antibiotic treatment (18), ensuring there was not already partial susceptibility to *C. difficile* before treatment. The additional controls in our study allowed us to assess the contribution of omeprazole alone to *C. difficile* susceptibility in mice.

Our study also extended previous work examining PPIs and *C. difficile* in mice by incorporating the contribution of the intestinal microbiota. We found that omeprazole had no significant impact on bacterial taxa within the murine intestinal microbiota over the 16-day experiment. In contrast to previous work with PPIs (15–17), omeprazole did not alter *C. difficile* colonization resistance in mice. 16S rRNA sequencing results suggested that *Streptococcus* and *Enterococcus* are rare genera in our C57BL/6 mouse colony. These two genera could be important contributors to the associations between PPIs and CDIs in humans, and could be a contributing factor to our observation that PPI treatment had no effect on *C. difficile* colonization in our CDI mouse model. While the intestinal microbiomes of both humans and mice are dominated by the *Bacteroidetes* and *Firmicutes* phyla, there are significant differences in the relative abundances of genera that are present and some genera are unique to each mammal (19), differences that may partly explain our results. Gastrointestinal physiological differences, particularly the higher stomach pH in mice (pH 3 to 4) compared to humans (pH 1) (19) could also explain why omeprazole had a limited impact on the murine microbiome. The microbiota and physiological differences between humans and mice may limit the usefulness of employing mouse models to study the impact of PPIs on the microbiota and CDIs.

Beyond microbiome differences, factors such as age, body mass index, comorbidities, and use of other medications in human studies may also be contributing to the association between PPIs and CDI incidence or recurrence. The type of *C. difficile* strain type could also be an important contributing factor; however, our study was limited in that we tested only *C. difficile* strain 630 (ribotype 012). This study addressed the impact of PPIs with or without antibiotics on a murine model of CDI and found that PPIs did not promote *C. difficile* colonization. The epidemiological evidence linking PPIs to CDIs is primarily from observational studies, which makes determining causality and whether other risk factors play a role challenging (20). Future studies are needed to determine whether age, other comorbidities, and bacterial strains that are less common in mice can increase the risk of CDIs or recurrent CDIs when combined with PPI treatment.

Experimental procedures. (i) Animals. All mouse experiments were performed with 7- to 12-week-old C57BL/6 male and female mice. Each experimental group of mice was split between two cages with two or three mice housed per cage and male and female mice housed separately. All animal experiments were approved by the University of Michigan Animal Care and Use Committee (IACUC) under protocol number PRO00006983.

(ii) Drug treatments. Omeprazole (Sigma-Aldrich) was prepared in a vehicle solution of 40% polyethylene glycol 400 (Sigma-Aldrich) in phosphate-buffered saline (PBS). Omeprazole was prepared from 20 mg/ml frozen aliquots and diluted to 8 mg/ml prior to gavage. All mice received 40 mg of omeprazole per kg of body weight (a dose previously used in mouse experiments [17]) or vehicle solution once per day through the duration of the experiment with treatment starting 7 days before *C. difficile* challenge (Fig. 1A). Although the omeprazole dose administered to mice is higher than the recommended dose for humans, omeprazole has a shorter half-life in mice compared to humans (21) and lacks an enteric coating (22). One day prior to *C. difficile* challenge, two groups of mice received an intraperitoneal injection of 10 mg/kg clindamycin or sterile saline vehicle (11). All drugs were filter sterilized through a 0.22- μ m syringe filter before administration to animals.

(iii) *C. difficile* infection model. Mice were challenged with *C. difficile* strain 630 7 days after the start of omeprazole treatment and 1 day after clindamycin treatment. Mice were challenged with 10^3 spores in ultrapure distilled water as described previously (11). Stool samples were collected for 16S rRNA sequencing or *C. difficile* CFU quantification throughout the duration of the experiments at the indicated time points (Fig. 1A). Samples for 16S rRNA sequencing were flash frozen in liquid nitrogen and stored at -80°C until DNA extraction, while samples for CFU quantification were transferred into an anaerobic chamber and serially diluted in PBS. Diluted samples were plated on TCCFA (taurocholate- cycloserine-cefoxitin-fructose agar) plates and incubated at 37°C for 24 h under anaerobic conditions to quantify *C. difficile* CFU.

(iv) 16S rRNA gene sequencing. DNA for 16S rRNA gene sequencing was extracted from 10 to 50 mg fecal pellet from each mouse using the DNeasy Powersoil HTP 96 kit (Qiagen) and an EpMotion 5075 automated pipetting system (Eppendorf). The 16S rRNA sequencing library was prepared as described previously (23). In brief, the ZymoBIOMICS microbial community DNA standard (Zymo, CA, USA) was used as a mock community (24), and water was used as a negative control. The V4 hypervariable region of the 16S rRNA gene was amplified with Accuprime Pfx DNA polymerase (Thermo Fisher Scientific) using previously described custom barcoded primers (23). The 16S rRNA amplicon library was sequenced with the MiSeq system (Illumina). Amplicons were cleaned up and normalized with the SequelPrep normalization plate kit (Thermo Fisher Scientific), and pooled amplicons were quantified with the KAPA library quantification kit (KAPA Biosystems).

(v) 16S rRNA gene sequence analysis. mothur (v1.40.5) was used for all sequence processing steps (25) using a previously published protocol (23). In brief, forward and reverse reads for each sample were combined, and low-quality sequences and chimeras were removed. Duplicate sequences were merged, before taxonomy assignment using a modified version (v16) of the Ribosomal Database Project reference database (v11.5) with an 80% cutoff. Operational taxonomic units (OTUs) were assigned with the optclust clustering algorithm using a 97% similarity threshold. To adjust for uneven sequencing across samples, all samples were rarefied to 3,000 sequences 1,000 times. PCoAs were generated based on Bray-Curtis distance. R (v.3.5.1) was used to generate figures and perform statistical analysis.

(vi) Statistical analysis. To test for differences in relative abundances in families and genera across our three different treatment groups at different time points (clindamycin, clindamycin plus omeprazole, and omeprazole on days -7 , 0 , 2 , and 9) or within the omeprazole treatment group across three time points (days -7 , 0 , and 9), we

used a Kruskal-Wallis test with a Benjamini-Hochberg correction for multiple comparisons.

Code availability. The code for all sequence processing and analysis steps as well as a Rmarkdown version of this manuscript is available at https://github.com/SchlossLab/Tomkovich_PPI_mSphere_2019.

Data availability. The 16S rRNA sequencing data have been deposited in the NCBI Sequence Read Archive (accession no. [PRJNA554866](https://www.ncbi.nlm.nih.gov/submit/sra/study/PRJNA554866)).

SUPPLEMENTAL MATERIAL

Supplemental material for this article may be found at <https://doi.org/10.1128/mSphere.00693-19>.

FIG S1, PDF file, 0.4 MB.

FIG S2, PDF file, 0.2 MB.

ACKNOWLEDGMENTS

This research was supported by NIH grant U01AI124255.

We thank the Unit for Laboratory Animal Medicine at the University of Michigan for maintaining our mouse colony and providing the infrastructure and support for performing our mouse experiments. We also thank members of the Schloss lab for helpful discussions throughout the process of designing the experiment, analyzing the results, crafting the figures, and writing drafts of the article.

REFERENCES

1. Schubert AM, Sinani H, Schloss PD. 2015. Antibiotic-induced alterations of the murine gut microbiota and subsequent effects on colonization resistance against *Clostridium difficile*. *mBio* 6:e00974-15. <https://doi.org/10.1128/mBio.00974-15>.
2. Tariq R, Singh S, Gupta A, Pardi DS, Khanna S. 2017. Association of gastric acid suppression with recurrent *Clostridium difficile* infection: a systematic review and meta-analysis. *JAMA Intern Med* 177:784–791. <https://doi.org/10.1001/jamainternmed.2017.0212>.
3. Nehra AK, Alexander JA, Loftus CG, Nehra V. 2018. Proton pump inhibitors: review of emerging concerns. *Mayo Clin Proc* 93:240–246. <https://doi.org/10.1016/j.mayocp.2017.10.022>.
4. Naito Y, Kashiwagi K, Takagi T, Andoh A, Inoue R. 2018. Intestinal dysbiosis secondary to proton-pump inhibitor use. *Digestion* 97:195–204. <https://doi.org/10.1159/000481813>.
5. Elias E, Targownik LE. 2019. The clinician's guide to proton pump inhibitor related adverse events. *Drugs* 79:715–731. <https://doi.org/10.1007/s40265-019-01110-3>.
6. Imhann F, Vila AV, Bonder MJ, Manosalva AGL, Koonen DP, Fu J, Wijmenga C, Zhernakova A, Weersma RK. 2017. The influence of proton pump inhibitors and other commonly used medication on the gut microbiota. *Gut Microbes* 8:351–358. <https://doi.org/10.1080/19490976.2017.1284732>.
7. Imhann F, Bonder MJ, Vila AV, Fu J, Mujagic Z, Vork L, Tigchelaar EF, Jankipersadsing SA, Cenit MC, Harmsen HJM, Dijkstra G, Franke L, Xavier RJ, Jonkers D, Wijmenga C, Weersma RK, Zhernakova A. 2016. Proton pump inhibitors affect the gut microbiome. *Gut* 65:740–748. <https://doi.org/10.1136/gutjnl-2015-310376>.
8. Freedberg DE, Toussaint NC, Chen SP, Ratner AJ, Whittier S, Wang TC, Wang HH, Abrams JA. 2015. Proton pump inhibitors alter specific taxa in the human gastrointestinal microbiome: a crossover trial. *Gastroenterology* 149:883–885.e9. <https://doi.org/10.1053/j.gastro.2015.06.043>.
9. Maier L, Pruteanu M, Kuhn M, Zeller G, Telzerow A, Anderson EE, Brochado AR, Fernandez KC, Dose H, Mori H, Patil KR, Bork P, Typas A. 2018. Extensive impact of non-antibiotic drugs on human gut bacteria. *Nature* 555:623–628. <https://doi.org/10.1038/nature25979>.
10. Schubert AM, Rogers MAM, Ring C, Mogle J, Petrosino JP, Young VB, Aronoff DM, Schloss PD. 2014. Microbiome data distinguish patients with *Clostridium difficile* infection and non-*C. difficile*-associated diarrhea from healthy controls. *mBio* 5:e01021-14. <https://doi.org/10.1128/mBio.01021-14>.
11. Jenior ML, Leslie JL, Young VB, Schloss PD. 2018. *Clostridium difficile* alters the structure and metabolism of distinct cecal microbiomes during initial infection to promote sustained colonization. *mSphere* 3:e00261-18. <https://doi.org/10.1128/mSphere.00261-18>.
12. Yasutomi E, Hoshi N, Adachi S, Otsuka T, Kong L, Ku Y, Yamairi H, Inoue J, Ishida T, Watanabe D, Ooi M, Yoshida M, Tsukimi T, Fukuda S, Azuma T. 2018. Proton pump inhibitors increase the susceptibility of mice to oral infection with enteropathogenic bacteria. *Dig Dis Sci* 63:881–889. <https://doi.org/10.1007/s10620-017-4905-3>.
13. Schmid SM, Suchodolski JS, Price JM, Tolbert MK. 2018. Omeprazole minimally alters the fecal microbial community in six cats: a pilot study. *Front Vet Sci* 5:79. <https://doi.org/10.3389/fvets.2018.00079>.
14. Castellani C, Singer G, Kashofer K, Huber-Zeyringer A, Flucher C, Kaiser M, Till H. 2017. The influence of proton pump inhibitors on the fecal microbiome of infants with gastroesophageal reflux—a prospective longitudinal interventional study. *Front Cell Infect Microbiol* 7:444. <https://doi.org/10.3389/fcimb.2017.00444>.
15. Kaur S, Vaishnavi C, Prasad KK, Ray P, Kochhar R. 2007. Comparative role of antibiotic and proton pump inhibitor in experimental *Clostridium difficile* infection in mice. *Microbiol Immunol* 51:1209–1214. <https://doi.org/10.1111/j.1348-0421.2007.tb04016.x>.
16. Kaur S, Vaishnavi C, Prasad KK, Ray P, Kochhar R. 2011. Effect of Lactobacillus acidophilus & epidermal growth factor on experimentally induced *Clostridium difficile* infection. *Indian J Med Res* 133:434–441.
17. Hung Y-P, Ko W-C, Chou P-H, Chen Y-H, Lin H-J, Liu Y-H, Tsai H-W, Lee J-C, Tsai P-J. 2015. Proton-pump inhibitor exposure aggravates *Clostridium difficile*-associated colitis: evidence from a mouse model. *J Infect Dis* 212:654–663. <https://doi.org/10.1093/infdis/jiv184>.
18. Jenior ML, Leslie JL, Young VB, Schloss PD. 2017. *Clostridium difficile* colonizes alternative nutrient niches during infection across distinct murine gut microbiomes. *mSystems* 2:e00063-17. <https://doi.org/10.1128/mSystems.00063-17>.
19. Hugenholtz F, de Vos WM. 2018. Mouse models for human intestinal microbiota research: a critical evaluation. *Cell Mol Life Sci* 75:149–160. <https://doi.org/10.1007/s00018-017-2693-8>.
20. Eze P, Balsells E, Kyaw MH, Nair H. 2017. Risk factors for *Clostridium difficile* infections – an overview of the evidence base and challenges in data synthesis. *J Glob Health* 7:e010417. <https://doi.org/10.7189/jogh.07.010417>.
21. Regårdh C-G, Gabrielsson M, Hoffman K-J, Löfberg I, Skånberg I. 1985. Pharmacokinetics and metabolism of omeprazole in animals and man – an overview. *Scand J Gastroenterol* 20:79–94. <https://doi.org/10.3109/00365528509095821>.
22. Llorente C, Jepsen P, Inamine T, Wang L, Bluemel S, Wang HJ, Loomba R, Bajaj JS, Schubert ML, Sikaroodi M, Gillevet PM, Xu J, Kisseleva T, Ho

- SB, DePew J, Du X, Sørensen HT, Vilstrup H, Nelson KE, Brenner DA, Fouts DE, Schnabl B. 2017. Gastric acid suppression promotes alcoholic liver disease by inducing overgrowth of intestinal enterococcus. *Nat Commun* 8:837. <https://doi.org/10.1038/s41467-017-01779-8>.
23. Kozich JJ, Westcott SL, Baxter NT, Highlander SK, Schloss PD. 2013. Development of a dual-index sequencing strategy and curation pipeline for analyzing amplicon sequence data on the MiSeq Illumina sequencing platform. *Appl Environ Microbiol* 79:5112–5120. <https://doi.org/10.1128/AEM.01043-13>.
24. Sze MA, Schloss PD. 2019. The impact of DNA polymerase and number of rounds of amplification in PCR on 16S rRNA gene sequence data. *mSphere* 4:e00163-19. <https://doi.org/10.1128/mSphere.00163-19>.
25. Schloss PD, Westcott SL, Ryabin T, Hall JR, Hartmann M, Hollister EB, Lesniewski RA, Oakley BB, Parks DH, Robinson CJ, Sahl JW, Stres B, Thallinger GG, Horn DJV, Weber CF. 2009. Introducing mothur: open-source, platform-independent, community-supported software for describing and comparing microbial communities. *Appl Environ Microbiol* 75:7537–7541. <https://doi.org/10.1128/AEM.01541-09>.